

Original Research Article

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Evaluation of Phenolics and Antioxidant Enzyme Systems for *Phytophthora* Blight in Resistant and Susceptible Variety of Sesame (*Sesamum indicum* L.)

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ABSTRACT

The present experiment was conducted with an objective to evaluate changes in total phenol and antioxidant enzymes in resistance and susceptible cultivars of sesames against *Phytophthora* blight. Among the sesame cultivars, root of resistant cultivar G. Til-10 showed the highest amount of total phenol content (0.846 mg.g⁻¹Fr.Wt.), while infected leaf tissue of susceptible cultivar G. Til-4 contained the lowest value (0.418 mg.g⁻¹Fr.Wt.). The activity of ROS scavenging enzymes superoxide dismutase (SOD), Guaiacol peroxidase (GPX), Ascorbate peroxidase (APX) and catalase (CAT) was remained higher in infected root and leaf tissue of the susceptible cultivar G. Til-4 compared to resistant genotypes. The activity of superoxide dismutase was increased an average 1-2 fold in susceptible genotype from pre-infection to infection stage. In case of GPX activity the activity continuously increased in infected tissue from pre-infection to post-infection stage with respect to non-infected tissue. The higher activity was recorded in susceptible cultivar G. Til-4. Same trend of result observed in ascorbate peroxidase activity. In case of polyphenol oxidase, highest activity was noted in infected leaf and root tissue of susceptible (G. Til-4) cultivar. The activity of catalase showed increased in activity of infected tissue compared to non-infected.

Keywords

Sesame, *Phytophthora* blight, Superoxide dismutase (SOD), Guaiacol peroxidase (GPX), Ascorbate peroxidase (APX), Catalase (CAT).

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Introduction

The sesame belongs to the family *Pedaliaceae* and genus *Sesamum* is a self-pollinated annual herb with a basic chromosome number of 2n=26. Sesame has important agricultural attributes: it is adapted to tropical and temperate conditions, grows well on stored soil moisture with minimal irrigation or rainfall can produce good yields under high temperatures, and its seed has a high nutritional value (Bennet, 2011).

It is also grown in all the crop growing seasons viz, *kharif*, late *kharif*, *rabi* and summer. It is grown in more than one season in some part and in different seasons in other

parts of the country. The main reason for low productivity in sesame is the adverse ecologies in which it is cropped and its vulnerability to abiotic and biotic stresses. *Phytophthora* blight is one of the biotic factors which adversely affect crops productivity of sesame. The differential reaction related to total phenol content and antioxidative enzymes for resistance and susceptible cultivars were observed when infected with pathogen. Thus the present experiment was planned to evaluate changes in total phenol and antioxidant enzymes in resistance and susceptible cultivars of sesames against *Phytophthora* blight.

Materials and Methods

Earthen pots (Approximately 18 cm diameter) were used for pot culture studies. Pots were sterilized with 5 per cent formaldehyde solution.

Field soil with farm yard manure (FYM) and sand were mixed in the proportion of 1:2 and sterilized in autoclave at 1:036kg/cm² for one hour for three consecutive days. Previously incubated flask with *Phytophthora* pathogen was then added to the soil in the proportion of 1:10 (inoculums and sterilized soil).

Diseases incidence

Diseases incidence of sesame cultivars were measured under pathogen infested soil culture.

Earthen pots were used for preparation of pathogen inoculum of approximately 4 x 10⁵ cfu g⁻¹ was mixed with soil three days prior to seed sowing.

Percentage of diseases incidence was recorded at different time interval based on pathogen infection on seedling (Akter *et al.*, 2007).

$$\text{Disease incidence (\%)} = \frac{\text{No. of disease plants}}{\text{Total No. of plants}} \times 100$$

Stage of infection (Sampling interval)

Sesame root and leaf tissues were collected at different stages of disease development. Different stages of infection were categorized as discussed by Dubey *et al.*, (2011).

The days after sowing corresponding to the stage of disease along with disease symptoms are given below.

Antioxidant enzyme assay

Sample extraction

Extract for determination of superoxide dismutase (SOD), guaiacol peroxidase (GPX), ascorbate peroxidase (APX), Polyphenol oxidase (PPO) and catalase (CAT), activities were prepared from 0.5 g of leaves and 0.3 g root tissue homogenized with a pre-chilled mortar and pestle under ice cold condition in 2 ml and 1 ml of extraction buffer respectively, containing 50 mM sodium phosphate buffer (pH 7.4). The homogenates were centrifuged at 10,000 rpm for 20 minutes and the supernatant was used for the assay.

Superoxide dismutase (SOD)

Total SOD (EC 1.15.1.1) activity was measured spectrophotometrically based on inhibition in the photochemical reduction of nitroblue tetrazolium (NBT). The 3 ml reaction mixture contained 50 mM sodium phosphate buffer (pH 7.8) 13 mM methionine, 75 μM NBT, 2 μM riboflavin, 0.1 mM EDTA and 0.05 ml enzyme extract, riboflavin was added last (Van Rossun *et al.*, 1997).

Test tubes were shaken and placed 30 cm below from a light blank consisting of four 15-w fluorescent lamps. The reaction was allowed to run for 10 minutes and stopped by switching the light off.

The photoreduction in NBT was measured as increase in absorbance at 560 nm. Blanks and controls were run the same way but without illumination and enzyme, respectively. One unit of SOD was defined as the quantity of enzyme required to inhibit the reduction of NBT by 50% in a reaction mixture. Enzyme unit of SOD was calculated according to formula given by (Constantine and Stanley, 1977).

Guaiacol peroxidase (GPX)

GPX (EC 1.11.1.7) activity was determined in the homogenates by measuring the increase in absorption at 470 nm due to the formation of tetraguaiacol ($\epsilon = 26.6 \text{ mM}^{-1} \text{ cm}^{-1}$) in a reaction mixture containing 50 mM sodium phosphate buffer pH 7.0, 0.1 mM EDTA, 25 μl enzyme extract, 10 mM guaiacol and 10 mM H_2O_2 (Costa *et al.*, 2002).

Ascorbate peroxidase (APX)

APX (EC 1.11.1.11) activity was measured immediately in fresh extract and was assayed as described by Nakano and Asada (1981). 3 ml reaction mixture containing 50 mM sodium phosphate buffer pH 7.0, 0.1 mM H_2O_2 , 0.5 mM ascorbic acid, 0.1 mM EDTA and 0.1 ml enzyme extract. The hydrogen peroxide dependent oxidation of ascorbate was followed by a decrease in the absorbance at 290 nm ($\epsilon = 2.8 \text{ mM}^{-1} \text{ cm}^{-1}$).

Polyphenol oxidase (PPO)

The PPO (EC 1.14.18.1) activity was measured using reaction mixture contained 2.9 ml of catechol (10mM catechol in 10 mM phosphate buffer, pH 6.5 and reaction was initiated by the addition of 100 μl of enzyme extract. The changes in the colour due to the oxidized catechol were read at 490 nm for one minute at an interval of 15 second. Blank was carried out without substrate. The enzyme activity was expressed $\text{U} \cdot \text{mg}^{-1} \text{ protein}$ and unit activity was defined as $\Delta \text{OD} \cdot \text{min}^{-1} \cdot \text{g}^{-1} \text{ Fr. Wt.}$ tissues (Malik and Singh, 1980).

Results and Discussion

Percent disease incidence

The sesame cultivars were grown in normal condition and blight sick condition. The percent disease incidence was based on blight

symptoms of sesame plants at up to 24 days after sowing (DAS) in a plot was recorded and showed statistically significant differences. The value in percent showed the tolerance or susceptibility towards disease incidence (Table 1).

The percent disease incidence was significantly varied from 4.46% to 99.84% with the advancement of disease (Table 1). The cultivars G. Til-10 showed less disease incidence (40.12 %) at post-infection stage (24 DAS), which was indicating tolerance to blight. The disease incidence was significantly increased to the tune of 15.60% to 99.84% in which G. Til-4 showed at post-infection stage (24 DAS).

Total phenol content

Total phenol content in root and leaf tissues of two cultivars grown in both normal (healthy) pot and from sick pot (diseased plants) obtained at three (S1, S2 and S3) disease development stages were recorded (Table 2). Among the different disease stages, phenol content significantly varied from 0.216 to 0.846 $\text{mg} \cdot \text{g}^{-1} \text{ Fr. Wt.}$. The phenol content significantly increased as development of different disease stages, from pre-infection to post-infection (S1 to S3). The root and leaf tissues obtained from infected plants revealed higher amount of total phenol content as compared to tissue in control plants. The mean effect of root tissue of resistance G. Til-10 was found highest (0.676 $\text{mg} \cdot \text{g}^{-1} \text{ Fr. Wt.}$) and lowest phenol content was observed in leaf tissue of susceptible cultivar G. Til-4 (0.310 $\text{mg} \cdot \text{g}^{-1} \text{ Fr. Wt.}$).

Irrespective of stages, infected root tissue of resistant G. Til-10 cultivar showed 1.42 fold increased in phenol content as compared to pre-infection stage. The minimal phenol content (0.216 $\text{mg} \cdot \text{g}^{-1} \text{ Fr. Wt.}$) was examined in leaf tissue of susceptible G. Til-4 at all

three stages of disease development. The combined effect of genotypes and disease development stages showed significant differences. Among both cultivars, infected root tissue of resistant cultivar G. Til-10 (0.846 mg.g⁻¹ Fr. Wt.) showed highest phenol content, compared with infected leaf tissue of G. Til-10 (0.699 mg.g⁻¹ Fr. Wt.).

Sharma *et al.*, (2011) studied the metabolic alterations in *Sesamum* after infection with *Macrophomina phaseolina* and *Fusarium oxysporum*. This accomplished individually by the levels of total phenolic compounds of one week old plants. In infected plants polyphenols along with salicylic acid (SA) considerably exceeded when compared to control plants.

Patel *et al.*, (2015) also reported the induction of phenol content in response to wilt pathogen infestation in pignonpea crop. Kandoliya and Vakharia (2013) also observe the higher phenol content in resistant variety of chickpea compared to susceptible variety in normal and infected plants of chickpea. This revealed inherent character of variety to cope with infection process.

Superoxide dismutase (SOD)

Superoxide dismutase (SOD) activity was examined in root and leaf tissue of resistant and susceptible cultivar of sesame. Irrespective of disease development stages,

mean effect of resistant cultivars of sesame showed significant difference (Table 3).

The mean SOD activity was found higher in root tissue of susceptible G. Til-4 (34.508 U.mg⁻¹ Fr.Wt.) cultivar followed by root tissue of resistant G. Til-10 (33.283 U.mg⁻¹ Fr.Wt.) cultivar). Among these cultivars, susceptible G. Til-4 (51.333 U.mg⁻¹ Fr.Wt.) cultivar showed highest activity at infection stage (S2). During stage of post-infection (S3) infected root tissue of resistant G. Til-10 cultivar showed 1.55 fold highest activity compared to non-infected and 1.53 fold highest activity compared to S1 stage. In infected root tissue of susceptible root tissue of G. Til-4 cultivar showed 1.18 fold higher activity compared to non-infected. Overall, Superoxide dismutase activity increased at S1 to S3 stage of disease development in resistant cultivar was found, whereas in susceptible cultivar there were slight decline in activity was found. SOD activity of infected root tissues were increased fast as compared to infected leaf tissues. Rajab *et al.*, (2009) examined the sesamum – *Fusarium* culture exudates interaction was accompanied by substantial increase in oxidative stress, probably as direct consequences of a progressive decline in the enzymatic system responsible for catabolism of active oxygen species. On the basis of quantitative data, among the enzymes tested, changes in SOD activity seemed to be related with the interaction of *Sesamum* × *Fusarium*.

Stage of infection (Sampling interval)			
Factor	Stage of disease	Days after sowing (DAS)	Visible symptoms of treatments grown in different pots
S ₁	Pre-inflectional stage	10	Normal, leaves were healthy and green in all treatments
S ₂	Disease inflectional stage	17	Brown spots on leaves in some of treatments.
S ₃	Post inflectional stage	22	Leaves shriveled and root gets brown, drooping of whole plants but not in all treatments.

Table.1 Percent (%) disease incidence at different growth periods (DAS) of seedlings

Sr. No.	Cultivars	% Disease infestation								
		16 DAS	17 DAS	18 DAS	19 DAS	20 DAS	21 DAS	22 DAS	23 DAS	24 DAS
1	G. Til-10	4.46	19.72	20.30	29.57	29.42	32.60	33.40	35.78	40.12
2	G. Til-4	15.60	21.75	35.37	47.56	55.36	61.23	72.42	89.50	99.84

Table.2 Total phenol content (mg.g⁻¹ Fr. Wt.) of sesame cultivars at three different disease stages

Sr. No.	Variety	Stage- 1		Stage- 2		Stage- 3		Mean (Vx)
		Non Infected	Infected	Non Infected	Infected	Non Infected	Infected	
1	G.Til-10 root	0.515	0.592	0.657	0.752	0.693	0.846	0.676
2	G.Till-10 leaf	0.430	0.490	0.513	0.590	0.556	0.699	0.546
3	G.Till-4 root	0.279	0.322	0.479	0.481	0.393	0.515	0.412
4	G.Till-4 leaf	0.216	0.297	0.276	0.351	0.301	0.418	0.310
	Mean (Sx)	0.360	0.425	0.481	0.544	0.486	0.620	
		S.Em ±		CD at 5 %		CV %		
Vx		0.0025		0.0071		2.17		
Sx		0.003		0.0085				
Vx X Sx		0.0061		0.0174				

Table.3 Superoxide dismutase activity (U.mg⁻¹ Fr. Wt.) at different disease development stages of *Phytophthora* blight in sesamum

Sr. No.	Variety	Stage- 1		Stage- 2		Stage- 3		Mean (Vx)
		Non Infected	Infected	Non Infected	Infected	Non Infected	Infected	
1	G.Til-10 root	26.323	27.453	36.443	40.153	27.083	42.243	33.283
2	G.Til-10 leaf	16.043	22.433	32.903	36.863	38.503	39.943	31.115
3	G.Til-4 root	21.833	44.463	33.143	51.333	25.723	30.553	34.508
4	G.Til-4 leaf	30.467	39.323	31.133	46.803	28.023	28.583	34.056
	Mean (Sx)	23.667	33.418	33.406	43.788	29.833	35.331	
		S.Em ±		CD at 5 %		CV %		
Vx		0.3795		1.0802		4.84		
Sx		0.4648		1.3229				
Vx X Sx		0.9295		2.6456				

$U = \Delta OD.min^{-1}.ml^{-1}$

Table.4 Guaiacol peroxidase activity (U.mg⁻¹ Fr. Wt.) at different disease development stages of *Phytophthora* blight in sesame

Sr. No.	Variety	Stage- 1		Stage- 2		Stage- 3		Mean (Vx)
		Non Infected	Infected	Non Infected	Infected	Non Infected	Infected	
1	G.Til-10 root	85.460	101.740	102.110	109.230	118.850	171.120	114.752
2	G.Till-10 leaf	85.380	90.610	95.120	108.990	107.490	140.790	104.730
3	G.Till-4 root	92.920	97.430	101.340	124.730	150.440	189.460	126.053
4	G.Till-4 leaf	72.240	77.480	128.780	148.970	189.010	211.750	138.038
	Mean (Sx)	84.000	91.815	106.838	122.980	141.448	178.280	
		S.Em ±			CD at 5 %		CV %	
	Vx	0.7095			2.0194		2.49	
	Sx	0.869			2.4734			
	Vx X Sx	1.7379			4.9465			

U = Δ OD.min⁻¹.ml⁻¹

Table.5 Ascorbate peroxidase activity (U.mg⁻¹ Fr. Wt.) at different disease development stages of *Phytophthora* blight in sesamum

Sr. No.	Variety	Stage- 1		Stage- 2		Stage- 3		Mean (Vx)
		Non Infected	Infected	Non Infected	Infected	Non Infected	Infected	
1	G.Til 10 root	2.013	2.573	3.773	7.933	3.853	9.453	4.933
2	G.Till-10 leaf	1.733	2.073	2.773	4.543	3.233	6.433	3.465
3	G.Till-4 root	4.173	7.853	12.653	14.413	17.133	22.183	13.068
4	G.Till-4 leaf	3.073	4.043	9.583	11.993	14.143	19.283	10.353
	Mean (Sx)	2.748	4.136	7.196	9.721	9.591	14.338	
		S.Em ±			CD at 5 %		CV %	
	Vx	0.0472			0.1343		2.52	
	Sx	0.0578			0.1645			
	Vx X Sx	0.1157			0.3293			

U = Δ OD.min⁻¹.ml⁻¹

Table.6 Polyphenol oxidase activity (U.mg⁻¹ Fr. Wt.) at different disease development stages of *Phytophthora* blight in sesamum

Sr. No.	Variety	Stage- 1		Stage- 2		Stage- 3		Mean (Vx)
		Non Infected	Infected	Non Infected	Infected	Non Infected	Infected	
1	G.Til-10 root	5.273	6.553	5.833	6.873	7.853	8.473	6.810
2	G.Til-10 leaf	5.313	6.153	5.393	7.873	6.433	8.593	6.627
3	G.Til-4 root	5.593	6.033	7.433	7.753	7.953	9.033	7.300
4	G.Til-4 leaf	5.773	6.203	6.993	7.793	7.933	8.983	7.280
	Mean (Sx)	5.488	6.236	6.413	7.573	7.543	8.771	
		S.Em ±		CD at 5 %		CV %		
	Vx	0.1226		0.3489		7.42		
	Sx	0.1501		0.4272				
	Vx X Sx	0.3002		0.8544				

U = Δ OD.min⁻¹.ml⁻¹

Guaiacol peroxidase (GPX)

GPX activity of root and leaf tissue was significantly elevated from pre-infection stage (S1) to post-infection stage (S3) with respect to infected to non-infected tissue. The mean GPX activity was found highest in leaf tissue of susceptible G. Til-4 cultivar (138.038 U.mg⁻¹ Fr.Wt.) followed by root tissue of susceptible G. Til-4 cultivar (126.053 U.mg⁻¹ Fr. Wt.). Among these cultivars, susceptible G. Til-4 cultivar showed 1.23 fold higher activity in their infected root tissue compared to non-infected root tissue followed by resistant G. Til-10 cultivar showed 1.06 fold higher activity in their infected root tissue compared to non-infected root tissue. During stage of post-infection (S3) infected root tissue of susceptible G. Til-4 cultivar showed 1.94 fold highest activity compared to S1 stage. In infected root tissue of resistant G. Til-10 cultivar showed 1.68 fold higher activity compared to S1 stage (Table 4). Overall, Guaiacol peroxidase activity significantly increased as increased the stage of disease development. An average, the root and leaf tissue of susceptible cultivar G.Til-4 shows

significantly higher activities as compared to resistance cultivar G. Til-10. Lubaina and Murugan, (2012) studied Induction of plant defence against pathogen attack is regulated by a complex network of different signals. The activity of antioxidant enzymes increased in response to pathogen inoculation. Increase in activity of GPX was insignificant after 24 hr post inoculation in the inoculated leaves. There was increase in GPX activity in root tissue of infected plant in the post infection stage from pre infection stage.

Ascorbate peroxidase (APX)

Ascorbate peroxidase (APX) activity was examined in root and leaf tissue of resistant and susceptible cultivars of sesame. The mean ascorbate peroxidase activity was found highest in root tissue of susceptible G. Til-4 (13.068 U.mg⁻¹ Fr.Wt.) cultivar followed by leaf tissue of resistant G. Til-10 (3.465 U.mg⁻¹ Fr.Wt.) cultivar (Table 5). The activity was increased from pre-infection to post-infection stage with higher elevated in infection tissue compared with non-infected tissue. Interaction effect

between cultivar and disease development stage show significance difference. Among these cultivars, susceptible G. Til-4 (22.183 U.mg⁻¹ Fr.Wt.) cultivar showed highest activity at S3 in root tissue followed by root tissue of resistant cultivar G. Til-10 (9.453 U.mg⁻¹ Fr.Wt.) (Table 5). The activity of G. Til-4 was elevated 1.29 fold higher in infected root tissue to non-infected at S3 stage. Overall, Ascorbate peroxidase activity significantly increased as increased the stage of disease development. Ascorbate peroxidase activity of root and leaf tissue at three different stages of infection is higher in susceptible cultivar G. Til-4 as compared to resistant cultivar G. Til-10. Kandoliya and Vakharia (2015) reported variable response of Ascorbat peroxidase activity in chickpea infected with wilt pathogen. Kadkhodaie *et al.*, (2013) showed that ascorbate peroxidase activity was increased with the level of infection increase. APX activity increased in post infection level as compared to control plant. There is a notable difference in susceptible and resistant genotype in APX enzyme activity. Furthermore, APX may play the role in-coordinating the expression of photo oxidative stress responsive genes APXs are involved not only in scavenging H₂O₂ but also in plant growth, development, lignifications, suberization, and cross-linking of cell wall compounds.

Polyphenol oxidase (PPO)

A progressive increase in the enzyme activity of the infected root and leaf tissue of resistant and susceptible cultivars were observed with the advancement of disease and growth of plants. During different infection stages, the mean effect showed highest activity in root tissue of susceptible cultivar G. Til-4 (7.300 U.mg⁻¹ Fr.Wt.) compare with root of resistant cultivar G. Til-10 (6.810 U.mg⁻¹ Fr.Wt.) (Table 6). Activity was significantly increased from pre-infection stage (S1) to post-infection stage (S3) with respect to non-infected tissue to infected tissue. The combination between cultivar and disease development stages showed significant difference. The post-infection stage (S3)

exhibited a continuous increase in the enzyme activities from the infection stage. Susceptible G. Til-4 cultivar showed 1.13 fold higher activity at S3 stage compared to non-infected root tissue, whereas resistant cultivar G. Til-10 showed 1.07 fold highest activity at compared to non-infected root tissue at S3 stage. Overall, an interaction between the varieties and disease development stages showed significantly higher PPO activities in susceptible as compared to resistant cultivar was found. Fallahpori *et al.*, (2013) observed, polyphenol oxidase enzyme was assayed as a resistance mechanism in resistant and susceptible germplasm. Evaluation of polyphenol oxidase activity in resistant and susceptible germplasms in 2, 4, 6, 8, 10 and 12 days after inoculation with *Fusarium oxysporum* f. sp. *Sesami* showed that polyphenol oxidase enzyme activity increases in susceptible germplasm with highest level in 4 days after inoculation. In susceptible germplasm, enzyme activity was increased slightly in low quantity. Kandoliya and Vakharia (2013) also observed the same response in chickpea during infection of wilt pathogen. In conclusion the results show that polyphenol oxidase activity play probable role in induction of plant resistance against outbreak of plant pathogens.

In conclusion among the sesamum cultivars, root of resistant cultivar G. Til-10 showed the highest amount of total phenol content. The activity of ROS scavenging enzymes superoxide dismutase (SOD), Guaiacol peroxidase (GPX) and Ascorbate peroxidase (APX) was remained higher in infected root and leaf tissue of the susceptible cultivar G. Til-4 compared to resistant genotypes. In case of polyphenol oxidase, highest activity was noted in infected leaf and root tissue of susceptible (G. Til-4) cultivar. This indicates inherent response of cultivars against pathogen in response to infection process.

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